

Theiler's murine encephalomyelitis virus

Host species

- natural hosts: wild mice [1], laboratory mice, [2] [3] [4], water, bank and meadow voles (family Microtinae) [5] [6]
- positive serological reactions in laboratory rats [7] [8]; virus might be related to TMEV-virus, only one report on clinical signs and lesions in rats (MHG-strain) [9], positive serological findings may indicate the presence of a yet uncharacterised virus (?rat cardiovirus?) [10]
- guinea pigs: the presence of antibodies to TMEV in guinea pigs suffering from lameness indicates that the causative agent of guinea pig lameness might be a cardiovirus [11]
- mice, rats, hamsters and cotton rats are susceptible to intracerebrally inoculated virus (GDVII strain) but not guinea pigs [12] [13].

Serological prevalence of TMEV in mouse and rat colonies:

Canada: 1980/1986: 40% of mouse colonies, 58% of rat colonies [8]

Germany: 1987-1988: 11% of mouse colonies, 20% of rat colonies [3]

France: 1996/1997: 26% of mouse colonies, 13% of rat colonies [2]

United States: 1996: 30% of mouse colonies, 5-10% of rat colonies [14]

Properties of the Virus

RNA-virus, family Picornaviridae, Genus *Cardiovirus*; all TMEV strains (see below) are of the same serotype and cross-neutralize with polyclonal antisera [15] [16].

- different subgroups exist:
 - subgroup TO (DA, BeAn 8386, WW, TO, Yale) may produce chronic persistent infection of the CNS, accompanied by demyelinating lesions of the spinal cord; small plaques in cell culture;
 - subgroup GDVII (FA, GDVII) produce acute fulminant encephalomyelitis; large plaques in cell culture [17] [18];
- virus can be stored for a long period at -60°C
- optimal stability of the virus in the vicinity of pH 8 and pH 3,3
- exposure to air has little influence on the stability of the virus
- TMEV is rapidly destroyed at temperatures above 50°C
- virus is completely inactivated by 1% H_2O_2 at 37°C and by 50% acetone or alcohol [19].

Strain susceptibility

different susceptibilities of various mouse strains after experimental intracerebral inoculation [17] [20]:

- high susceptibility: SJL/J, DBA/1, DBA/2, SWR, PL/J and NZW mice
- intermediate susceptibility: C3H, CBA, AKR, C57BR mice
- resistant: BALB/C substrains, C57BL/6, C57BL/10, C57/L, 129/Jm and H-2D(b) mice; resistance to DA-virus in H-2(b) mice maps to the H-2D gene and is associated with a potent antiviral cytotoxic T-lymphocyte response [21]
- with cyclophosphamide (cy), mice can be made susceptible but resistance was restored by adoptive transfer of splenic cells from non cy treated donors, only C57Bl/6 could not be made susceptible; high doses of gamma irradiation increase susceptibility of mice [20]

Organotropism

replication of the virus in gastrointestinal mucosa [22] [23]; natural infection rarely spreads from intestine to spinal cord or brain; macrophages are a reservoir of the virus (Da, To, WW, BeAn) [24] [25] as well as oligodendrocytes, astrocytes and microglia [26] [27]; placentas and fetuses (only in early gestation) can be infected [28];

Clinical disease

- **mice, natural infection, subgroup TO (DA, BeAn 8386, WW, TO, Yale):** in mice asymptomatic gastrointestinal infection (except immunodeficient mice [29]), the virus rarely spreads to the central nervous system [30] [13], symptoms are flaccid posterior paralysis and seldom anterior paralysis in mice that are otherwise clinically normal; incubation period: 7-30 days
- **mice, natural infection, subgroup GDVII (FA, GDVII):** encephalomyelitic form may be expressed clinically by excitability, circling, rolling, tremor and convulsions on noise stimulation (incubation time: 2-9 days); most of the infected mice die soon after onset of clinical signs [23]
- **Rats, MHG-virus strain:** case report of symptoms in 3 rats of a colony with symptoms like circling, incoordination, tremor, torticollis [9]
- **experimental infections in mice and rats (intracerebrally, intranasally or footpad inoculation): strains DA, BeAn, WW, TO, Yale:** wobbling gait, about 2 to 4 weeks p.i., followed by weakness of the posterior limbs, spastic paralysis, urinary incontinence and priapism [31] [32]; weanling rats die within 2-3 days without paralytic symptoms [12] strains FA, GDVII: hyperexcitability, circling, and flaccid paralysis which lead to death within one week [23] [33] [34]

Pathology

- **mice, natural infection, subgroup TO (DA, BeAn 8386, WW, TO, Yale):** non-suppurative encephalomyelitis with gliosis and necrosis of ventral horn neurons of the spinal cord and neuronal necrosis in posterior regions of the brain, satellitosis,

Cowdry type B intranuclear inclusion bodies in neurons are not a consistent feature of the disease [22], inflammation may persist for several months after necrosis subsides and is then accompanied by astrocytosis and focal mineralization [35]

- **mice, natural infection, subgroup GDVII (SCID-mice):** severe degeneration (often spongiform) and necrosis of neurones and glial cells of the ventral horns (lesser involvement of the dorsal horns of the spinal cord) [29]
- **experimental infections in mice (intracerebral inoculation):**
- **subgroup: DA, BeAn, WW, TO, Yale:** acute neuronal degenerative changes and microglial proliferation primarily in the spinal cord anterior horn, brain stem and thalamus and perivascular inflammation also in the spinal cord leptomeninges, followed after 1 month p.i. by persistent viral infection of the spinal cord (white matter) with varying degrees of chronic progressive demyelination and inflammation and remyelination after a few months (resembles multiple sclerosis in man [31] [12] [33]); Hydrocephalus and pachymeningitis in mice after inoculation of an DA virus variant (H101 virus), without viral persistence, no demyelination [36]•subgroup: FA, GDVII: acute polioencephalomyelitis with necrosis of ganglion cells and neuronophagia of hippocampus, cortex and spinal cord anterior horn and nonsuppurative inflammation, high apoptosis rate in neurons, little if any demyelination, no viral persistence in the CNS [37] [38] [39]

Morbidity and mortality

- **natural infection:** subgroup TO (DA, BeAn 8386, WW, TO, Yale): morbidity low, little or no mortality (except immunodeficient mice with high morbidity and mortality [29]); strains FA, GDVII: morbidity and mortality high
- **experimental infection:** (intracerebral inoculation): subgroup TO (DA, BeAn 8386, WW, TO, Yale): morbidity high, mortality low; strains FA, GDVII: morbidity high (100%), mortality high

Zoonotic potential

- none

Interference with research

- chronic CD4+ response which is initially directed at viral determinants but persists in the CNS and is directed against multiple myelin autoepitopes; T cell proliferative response in spleen [32]
- increase of CD4+ Th1-cells producing IFN-gamma (DA-strain) [40]
- high m-RNA expression of proinflammatory cytokines in brain and spinal cord of SJL/J mice beginning at day 5 post infection for tumour necrosis factor- α (TNF- α) and interferon- γ (INF- γ); with DA additionally for lymphotoxins LT- α and LT- β , and with GDVII additionally for interferon- β (INF- β) and interleukin-6 (IL-6) and high m-RNA chemokine expression after DA, GDVII and H101-virus infection for Rantes, monocyte chemoattractant protein-1 (MCP-1), IP-10 and macrophage inflammatory proteins (MIP-1 β , MIP-1 α , MIP-2) [41] [42] [43]
- Increase of INF- α and INF- β in SJL CD4^{-/-} mice (DA-strain) [44]

- Interleukin-1 receptor declines in hippocampus in susceptible strains (SJL/L) [45]
- high apoptosis rate in neurons in GDVII infected mice, in DA infected mice high apoptosis rate in oligodendrocytes [39]
- inhibition of alpha/beta interferon synthesis in infected L929 cells [46]
- restraint stress has an effect on infected (BeAn strain) animals (increased mortality, increased viral titres, decreased number of lymphocytes) [47]

References

- Lipton, H.L., et al., Serological evidence that *Mus musculus* is the natural host of Theiler's murine encephalomyelitis virus. *Virus Res*, 2001. 76(1): p. 79-86.
- Zenner, L. and J.P. Regnault, Ten-year long monitoring of laboratory mouse and rat colonies in French facilities: a retrospective study. *Lab Anim*, 2000. 34(1): p. 76-83.
- Kraft, V. and B. Meyer, Seromonitoring in small laboratory animal colonies. A five year survey: 1984-1988. *Z Versuchstierkd*, 1990. 33(1): p. 29-35.
- Miyata, H. and H. Sato, Theiler's murine encephalomyelitis virus--characterization of newly isolated viruses from Japanese mouse colonies. *Jikken Dobutsu*, 1990. 39(4): p. 539-48.
- Descoteaux, J.P. and S. Mihok, Serologic study on the prevalence of murine viruses in a population of wild meadow voles (*Microtus pennsylvanicus*). *J Wildl Dis*, 1986. 22(3): p. 314-9.
- Descoteaux, J.P., [Serological study of the incidence of murine viruses in a population of small wild rodents (*Microtus pennsylvanicus* Ord, 1815)]. *Rev Sci Tech*, 1992. 11(4): p. 1071-7.
- Hemelt, I.E., D.L. Huxsoll, and A.R. Warner, Jr., Comparison of MHG virus with mouse encephalomyelitis viruses. *Lab Anim Sci*, 1974. 24(3): p. 523-9.
- Lussier, G. and J.P. Descoteaux, Prevalence of natural virus infections in laboratory mice and rats used in Canada. *Lab Anim Sci*, 1986. 36(2): p. 145-8.
- Mc Connell, S.J., Huxsoll, D. L., Garner, F.M., Spertzel, R.O., Warner, A.R. and Yager, R.H., Isolation and characterization of a neurotropic agent (MHG virus) from adult rats. *Proceedings of the society for experimental biology and medicine*, 1964. 115: p. 362-267.
- Oshawa, K., Watanabe, Y., Miyata, H. and Sato, H., Genetic analysis of TMEV-like virus isolated from rats: nucleic characterization of 3D protein region. *ALAAS*, 1998.
- Hansen, A.K., P. Thomsen, and H.J. Jensen, A serological indication of the existence of a guinea pig poliovirus. *Lab Anim*, 1997. 31(3): p. 212-8.
- Downs, W.G., Mouse encephalomyelitis virus. *The mouse in biomedical research*, 1982. 2: p. 341-352.
- Thompson, R., Harrison, W.M., Meyer, F.P., A spontaneous epizootic of mouse encephalomyelitis. *Proceedings of the society for experimental biology and medicine*, 1951. 77: p. 262-266.
- Jacoby, R.O. and J.R. Lindsey, Risks of Infection among Laboratory Rats and Mice at Major Biomedical Research Institutions. *Ilar J*, 1998. 39(4): p. 266-271.
- Rozhon, E.J., H.L. Lipton, and F. Brown, Characterization of Theiler's murine encephalomyelitis virus RNA. *J Gen Virol*, 1982. 61(Pt 2): p. 157-65.

Ohara, Y. and R. Roos, The antibody response in Theiler's virus infection: new perspectives on multiple sclerosis. *Prog Med Virol*, 1987. 34: p. 156-79.

Lipton, H.L. and M.C. Dal Canto, The TO strains of Theiler's viruses cause "slow virus-like" infections in mice. *Ann Neurol*, 1979. 6(1): p. 25-8.

Yamada, M., A. Zurbriggen, and R.S. Fujinami, Pathogenesis of Theiler's murine encephalomyelitis virus. *Adv Virus Res*, 1991. 39: p. 291-320.

Theiler M, G.S., Encephalomyelitis of mice. I. Characteristics and pathogenesis of the virus. *Exp Med*, 1940a. 72: p. 49-67.

Dal Canto, M.C., R.W. Melvold, and B.S. Kim, A hybrid between a resistant and a susceptible strain of mouse alters the pattern of Theiler's murine encephalomyelitis virus-induced white matter disease and favors oligodendrocyte-mediated remyelination. *Mult Scler*, 1995. 1(2): p. 95-103.

Azoulay-Cayla, A., et al., Roles of the H-2D(b) and H-K(b) genes in resistance to persistent Theiler's murine encephalomyelitis virus infection of the central nervous system. *J Gen Virol*, 2001. 82(Pt 5): p. 1043-7.

Olitsky PK, S.R., Histopathology of CNS of mice infected with virus of Theiler's disease. *Proc Soc Exp Biol Med*, 1941. 47: p. 79-83.

Theiler, M., Gard, S., Encephalomyelitis of mice III: Epidemiology. *Journal of experimental medicine*, 1940b. 72: p. 79-90.

Dal Canto, M.C. and S.G. Rabinowitz, Experimental models of virus-induced demyelination of the central nervous system. *Ann Neurol*, 1982. 11(2): p. 109-27.

Clatch, R.J., et al., Theiler's murine encephalomyelitis virus (TMEV)-induced demyelinating disease in mice is influenced by the H-2D region: correlation with TEMV-specific delayed-type hypersensitivity. *J Immunol*, 1985. 135(2): p. 1408-14.

Aubert, C. and S. Ozden, Comparison of the sensitivities of ribonucleic acid and oligonucleotide probes for in situ detection of Theiler's virus mRNA. *J Histochem Cytochem*, 1993. 41(7): p. 1099-103.

Rodriguez, M., J.L. Leibowitz, and P.W. Lampert, Persistent infection of oligodendrocytes in Theiler's virus-induced encephalomyelitis. *Ann Neurol*, 1983. 13(4): p. 426-33.

Abzug, M.J., Identification of trophoblastic giant cells as the initial principal target of early gestational murine enterovirus infection. *Placenta*, 1993. 14(2): p. 137-48.

Rozengurt, N. and S. Sanchez, Vacuolar neuronal degeneration in the ventral horns of SCID mice in naturally occurring Theiler's encephalomyelitis. *J Comp Pathol*, 1992. 107(4): p. 389-98.

Theiler, M., Spontaneous encephalomyelitis of mice: a new virus disease. *Journal of experimental medicine*, 1937. 65: p. 705-19.

Lipton, H.L., Theiler's virus infection in mice: an unusual biphasic disease process leading to demyelination. *Infect Immun*, 1975. 11(5): p. 1147-55.

Dal Canto, M.C., et al., Theiler's Murine Encephalomyelitis Virus (TMEV)-Induced Demyelination: A Model for Human Multiple Sclerosis. *Methods*, 1996. 10(3): p. 453-61.

Rodriguez, M., M.L. Pierce, and E.A. Howie, Immune response gene products (Ia antigens) on glial and endothelial cells in virus-induced demyelination. *J Immunol*, 1987. 138(10): p. 3438-42.

Martinat, C., et al., The GDVII strain of Theiler's virus spreads via axonal transport. *J Virol*, 1999. 73(7): p. 6093-8.

Jacoby, R., Encephalomyelitis, Theiler's Virus, Mouse. *Nervous System, Monographs on Pathology of Laboratory Animals*, ed. U.M. T.C. Jones, R.D.Hunt. 1988: Springer-Verlag.

- Tsunoda, I., et al., Exacerbation of viral and autoimmune animal models for multiple sclerosis by bacterial DNA. *Brain Pathol*, 1999. 9(3): p. 481-93.
- Lipton, H.L., Persistent Theiler's murine encephalomyelitis virus infection in mice depends on plaque size. *J Gen Virol*, 1980. 46(1): p. 169-77.
- Obuchi, M. and Y. Ohara, Theiler's murine encephalomyelitis virus (TMEV): the role of a small out-of-frame protein in viral persistence and demyelination. *Jpn J Infect Dis*, 1999. 52(6): p. 228-33.
- Tsunoda, I., C.I. Kurtz, and R.S. Fujinami, Apoptosis in acute and chronic central nervous system disease induced by Theiler's murine encephalomyelitis virus. *Virology*, 1997. 228(2): p. 388-93.
- Monteyne, P., et al., The Th1/Th2 balance does not account for the difference of susceptibility of mouse strains to Theiler's virus persistent infection. *J Immunol*, 1999. 162(12): p. 7330-4.
- Theil, D.J., et al., Alterations in cytokine but not chemokine mRNA expression during three distinct Theiler's virus infections. *J Neuroimmunol*, 2000. 104(1): p. 22-30.
- Hoffman, L.M., et al., Central nervous system chemokine expression during Theiler's virus-induced demyelinating disease. *J Neurovirol*, 1999. 5(6): p. 635-42.
- Murray, P.D., et al., Biphasic and regionally-restricted chemokine expression in the central nervous system in the Theiler's virus model of multiple sclerosis. *J Neurovirol*, 2000. 6 Suppl 1: p. S44-52.
- Rodriguez, M., et al., The CD4-mediated immune response is critical in determining the outcome of infection using Theiler's viruses with VP1 capsid protein point mutations. *Virology*, 2000. 275(1): p. 9-19.
- Lledo, A., J. Borrell, and C. Guaza, Dexamethasone regulation of interleukin-1-receptors in the hippocampus of Theiler's virus-infected mice: effects on virus-mediated demyelination. *Eur J Pharmacol*, 1999. 372(1): p. 75-83.
- van Pesch, V., O. van Eyll, and T. Michiels, The leader protein of Theiler's virus inhibits immediate-early alpha/beta interferon production. *J Virol*, 2001. 75(17): p. 7811-7.
- Campbell, T., et al., The effects of restraint stress on the neuropathogenesis of Theiler's virus infection: I. Acute disease. *Brain Behav Immun*, 2001. 15(3): p. 235-54.

Author: Bettina Kränzlin, Klinikum Mannheim, Germany